Received: 02.05.2024 Accepted: 05.08.2024 Published: 11.10.2024 POLISH Journal *of* Radiology

Review paper

The peritumoral brain zone in glioblastoma: a review of the pretreatment approach

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Abstract

Glioblastomas are the most common and aggressive form of malignant primary brain tumors in adults. The standard treatment is surgical resection followed by radiotherapy and chemotherapy. Despite optimal treatment methods, the prognosis for patients remains poor.

Preoperative determination of glioblastoma margins remains beneficial for the complete removal of the tumor mass. Radiotherapy is essential for post-surgery treatment, but radioresistance is a significant challenge contributing to high mortality rates. Advanced imaging technologies are used to analyze the changes in the peritumoral brain zone (PTZ). Consequently, they may lead to the development of novel therapeutic options, especially targeting the marginal parts of a tumor, which could improve the prognosis of glioblastoma patients.

The clinical presentation of glioblastoma is heterogeneous and mostly depends on the location and size of a tumor. Glioblastomas are characterized by both intratumoral cellular heterogeneity and an extensive, diffuse infiltration into the normal tissue bordering a tumor called the PTZ. Neuroimaging techniques, such as diffusion-weighted imaging (DWI), diffusion tensor imaging (DTI), perfusion-weighted imaging (PWI), proton magnetic resonance spectroscopy (¹H MRS), and chemical exchange saturation transfer (CEST) are useful methods in the evaluation of the tumor infiltration and thus the resection margin.

Key words: glioblastoma, peritumoral brain zone, surgery, neuroimaging.

Introduction

Glioblastomas are the most common and aggressive form of malignant primary brain tumors in adults [1]. The annual incidence of glioblastoma is approximately 3.22 per 100,000 individuals. The overall survival (OS) rate remains low, despite optimal surgical and postoperative therapies. The five-year survival rate for glioblastoma is approximately 6.8% in the USA [2]. The treatment options depend on the tumor type and include a combination of neurosurgery, chemotherapy with temozolomide, and radiotherapy [3]. Recent data indicated that the median OS in the case of total resection was 15.5 months versus 11.7 months for individuals with subtotal resection [4]. However, the infiltrative tumor growth patterns and diffuse brain involvement restrict the possibility of complete resection of the tumor mass [5]. It is noteworthy that recurrence of glioblastoma is expected in 90% of individuals within 12 months after the diagnosis [6].

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Authors' contribution:

A Study design · B Data collection · C Statistical analysis · D Data interpretation · E Manuscript preparation · F Literature search · G Funds collection

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Radiological features of the tumor boundaries are identified by magnetic resonance imaging (MRI) features before and after application of a gadolinium-based contrast agent. Several studies have shown tumor cells infiltrating into the apparently normal adjacent brain tissue where the recurrence is expected to occur. Conventional MRI sequences might underestimate tumor spread due to the limitations in distinguishing purely vasogenic and glioma-infiltrated edema [7]. Recently, the MRI techniques have been refined and allow us to determine glioma spread, especially through chemical exchange saturation transfer (CEST), but also relaxometry, perfusion, and diffusion [5,6]. Thus, the peritumoral brain zone (PTZ) has become the subject of intensive investigations for accurate assessment of the tumor margin and the oncogenic potential of bordering tumor brain tissue and radiotherapy planning [8-10].

The present review aims to provide current data about the importance of the PTZ for more effective treatment and to improve the prognosis among patients diagnosed with glioblastoma.

Glioblastoma and peritumoral zone

Glioblastoma is a primary brain tumor arising from glial cells, categorized as grade 4 according to the definition provided by the World Health Organization (WHO). A novel approach to glioblastoma grading is based on genetic alterations according to molecular biomarkers. The presence of specified mutations provides genetic markers for gliomas, especially isocitrate dehydrogenase (IDH) mutation status, 1p/19q co-deletion, ATRX gene mutations, MGMT promoter methylation status, H3F3A alterations, loss of CDKN2A, EGFR amplification, chromosome 7 amplification and chromosome 10 deletion, and TERT promoter pathogenic variants [11]. In IDH-wildtype diffuse astrocytic gliomas, the presence of one of the specific genetic parameters, such as EGFR gene amplification, TERT promoter pathogenic variants, or a combination of chromosome 7 amplification and 10 deletions, classifies them as glioblastoma [12]. Glioblastoma is distinguished into four molecular subtypes: proneural, classical, neural, and mesenchymal. The tumor zone (TZ) in glioblastoma is characterized by an immunosuppressive microenvironment through the production of immunosuppressive enzymes and the occurrence of other immune mechanisms and immunosuppressive cell populations able to suppress T-cell functionality [13]. Glioblastomas are characterized by both intratumoral cellular heterogeneity and extensive, diffuse infiltration into the brain parenchyma. Morphologically, these tumors commonly exhibit features such as necrosis and microvascular hyperplasia [14].

The area of gadolinium enhancement on T1-weighted imaging (T1) has limited potential to precisely delineate the tumor mass, including tumor invasion into the surrounding normal-appearing brain tissue (Figure 1) [7]. The PTZ holds notable significance due to its pro-oncogenic potential, as tumor recurrence frequently manifests at the margin of the resection cavity [6]. Notably, the PTA shows strongly reduced immunosuppression in comparison to the TC. In the literature, there is no universally agreed upon definition of the PTZ, probably due to the limited understanding of this region.

Some authors defined this region as the non-enhancing brain area of several centimeters in width surrounding the contrast-enhancing region of the tumor [15]. Inoue *et al.* [16] investigated the relationship of tumor imaging volume between MRI and ¹¹C-methionine (Met)-positron

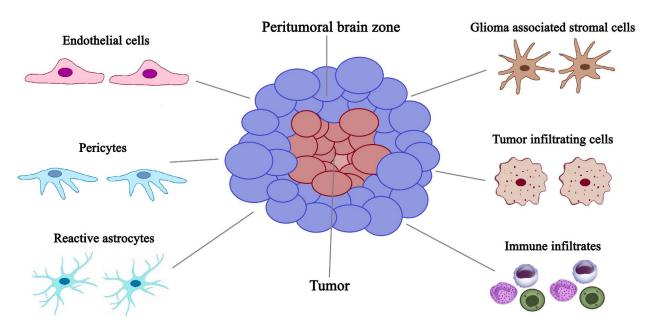


Figure 1. Graphic presentation of the cellular heterogeneity in the peritumoral brain zone (PTZ) that surrounds the glioblastoma

emission tomography (PET), and also the relationship between Met uptake index and tumor activity. This work revealed that 11C-Met-PET could delineate tumor extension more accurately than Gd-enhanced tumor mass on MRI. Considering the high infiltrative growth pattern of gliomas, Sebok et al. [17] characterized the extent of diffuse glioma tissue infiltration beyond the visible lesion (i.e., beyond the T1 contrast-enhanced lesion and/ or T2-weighted imaging (T2)/fluid-attenuated inversion recovery (FLAIR) – defined tumor border), with ¹⁸F-FET PET and functional MRI-based cerebrovascular reactivity (CVR) mapping. Volumes of interest starting from the tumor lesion outward up to 30 mm were created for a detailed peritumoral PET and MRI-CVR tissue analysis. The authors detected hypermetabolism and impaired cerebrovascular reactivity beyond the standard MRIdefined tumor border, suggesting active tumor infiltration in the peritumoral tissue. The hypermetabolism was still significantly present 6 mm beyond the visible glioma lesion, whereas significant MRI-CVR impairment was observed up to 12 mm. In this review, the PTZ is radiologically defined as the non-enhancing brain area of several centimeters in width surrounding the tumor mass.

Clinical presentation

The clinical presentation of glioblastoma is heterogeneous and mostly depends on the location and size of a tumor. Symptoms may develop abruptly, mimicking a stroke, or develop progressively over the years [18]. The common symptoms manifest in focal neurological deficits and increased intracranial pressure. Seizures and headaches, typically one-sided, are the most common initial symptoms in this group of patients. The pathophysiology of epilepsy arises from a combination of intrinsic epileptogenic properties of tumor tissue and the surrounding microenvironment as well as the disruption of nearby brain structures [19]. Focal neurological deficits occur in variable forms such as hemiparesis, aphasia, cognitive or behavioral changes, and vision or hearing problems.

Patients seek medical evaluation due to reported health issues and often undergo a computed tomography (CT) scan. When other potential causes are excluded and a tumor lesion is identified, diagnosis relies on MRI [20].

Magnetic resonance imaging

Neuroimaging of glioblastoma is commonly based on conventional MRI sequences including T1, T2, FLAIR, T2* gradient echo, and contrast-enhanced T1 images. However, the conventional MRI features have some limitations, for instance in distinguishing various glioma grades [6].

Glioblastomas demonstrate a variable signal on T1 and T2, heterogeneous contrast enhancement, irregular boundaries, as well as necrosis, hemorrhage and marked vasogenic edema [21,22]. The preoperative contrastenhanced MRI is a dedicated method to estimate the boundaries of glioblastoma, but the precise assessment of the PTZ is not a standard analysis. However, some studies have shown that tumor invasion into normal-appearing brain tissue may occur [5,10,23].

Typically, the PTZ shows hyperintensity on T2 and FLAIR images due to vasogenic edema driven by the disruption of the blood-brain barrier, increased capillary permeability, a pressure gradient from blood vessels to the extracellular environment, and the infiltration of proteins into the extracellular space. The non-enhancing regions of brain tissue bordering the tumor mass may show features that are not radiologically specific to tumor tissue and may be indistinguishable from vasogenic edema and other alterations [10,24,25]. Previous studies confirmed a combination of vasogenic edema and tumor infiltration in the PTZ [7,26]. Some tissue changes such as cell migration, tumor infiltration, neoangiogenesis, immune changes and edema formation are expected to coincide with certain functional and microstructural modifications that can be detected using advanced MRI techniques. Indeed, additional neuroimaging techniques such as diffusion-weighted imaging (DWI), diffusion tensor imaging (DTI), perfusion-weighted imaging (PWI), proton magnetic resonance spectroscopy (¹H MRS), and CEST could improve the pre-operative evaluation of glioblastoma [25,26].

Multidirectional DWI allows for the calculation of quantitative water diffusion maps, including the apparent diffusion coefficient (ADC), fractional anisotropy, mean diffusivity, radial diffusivity, and axial diffusivity [6,8]. Other promising methods in neurooncologic imaging include diffusion kurtosis imaging (DKI) and intravoxel incoherent motion (IVIM). IVIM has been used as an extension of DWI and ADC by using a spectrum of b values to assess tissue perfusion when contrast agents could not be safely administered. DKI is an advanced modality that extends conventional DTI by estimating the kurtosis of water diffusion based on a probability distribution function. However, the mentioned techniques are not widely adopted for routine clinical practice [27].

MRS provides tissue characterization, which is a clinical tool used to evaluate neurologic disorders through chemical shift imaging of hydrogen and other atoms, such as phosphorus, carbon, and sodium [28]. ¹H MRS is a valuable imaging tool for examining brain tumor metabolism by analyzing the presence of certain chemical compounds. In clinical practice, ¹H MRS detects metabolites such as N-acetylaspartate (NAA), choline (Cho), lactate (Lac), mobile lipids, creatine (Cr), glutamate (Glu), glutamine (Gln), glycine (Gly), myoinositol (mI), and glutathione (GSH). This tool, using sequences such as 3D echo planar spectroscopic imaging (3D-EPSI), allows for the mapping of metabolite alterations in different tumor regions [29]. In the literature, positive correlations between

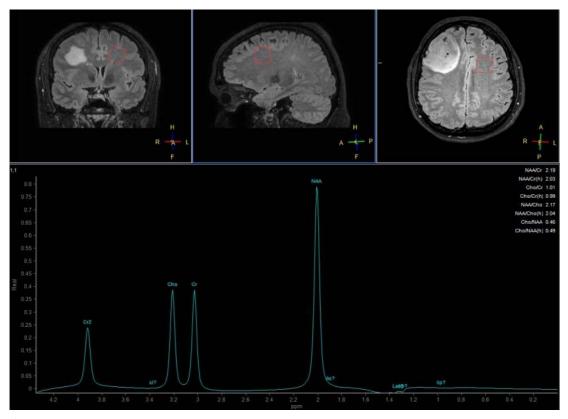


Figure 2. ¹H-MRS spectra showed a higher Cho/NAA ratio in the tumoral as well as peritumoral zone

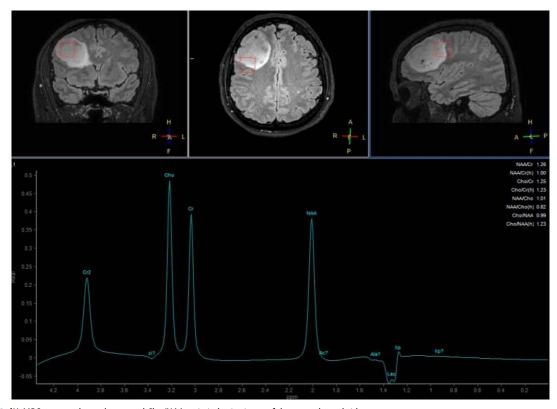


Figure 3. ¹H-MRS spectra showed a normal Cho/NAA ratio in brain tissue of the contralateral side

Cho/NAA ratios and quantitative measures of tumor infiltration have been reported, which means that whole-brain metabolic maps could be used for the detection of tumor borders (Figures 2 and 3). Considering the biochemical potential of assessment of various metabolic conditions, these techniques show promise in assessing tumor grade,

planning radiation therapy, and even distinguishing tissue changes between recurrent tumors versus radiation injury [29,30]. Unfortunately, MRS is limited by a considerable overlap between the spectroscopic appearance of different pathologies. MRS and other imaging modalities should be combined to precisely assess the abnormality [31].

Amide proton transfer (APT) CEST is a modern metabolic imaging method that generates image contrast by reflecting the associations of mobile proteins and peptides that resonate at 3.5 ppm downfield from water. The changes in the brain tumor microenvironment play crucial roles in understanding tumor development, growth and progression. APT-CEST imaging has shown potential to differentiate high- and low-grade gliomas and estimate the nature of lesions [32]. On the other hand, Akbari et al. observed a significant correlation between maps of acidity and hyperintense PTZ in T2 and FLAIR [33]. According to another study, higher CEST signals were observed from the PTZ in gliomas compared to contralateral brain regions and were associated with seizure activities [34]. These imaging techniques could lead to distinguishing between vasogenic edema and tumor infiltration (Figure 4). Importantly, CEST imaging is being studied for routine application in daily practice [35].

The integration of information from advanced neuroimaging techniques with conventional MRI sequences could improve pre-operative assessment and indicate regions at the highest risk of tumor recurrence.

Moreover, the importance of the peri-tumoral zone is evidenced by the numerous publications based on the analysis of radiomic features extracted from this area. The concept of radiomics involves extracting from large amounts of quantitative data from images. Such radiomic data describe the images mathematically, and their detailed analysis by machine learning models has a potential role in diagnosis and prognosis [36]. Some reports using radiomics demonstrate that, based on radiomic features of the peri-tumoral zone, it is possible to predict longversus short-term survival [37]. Other studies indicate that radiomic features of the peri-tumoral zone can be used to predict the site where glioblastoma recurrence will occur [38]. It is worth noting that radiomics, which has been applied in assessment of the peri-tumoral zone for a relatively short time, is already able to prove its heterogeneity.

Treatment

Patients with glioblastoma have a poor prognosis despite the current standard of care, including surgery, radiotherapy, and pharmacotherapy, typically chemotherapy with concomitant temozolomide [39]. The standard initial approach is surgical resection, which enables an accurate histological analysis, tumor genotyping, and a reduction in tumor mass. Radiotherapy with concomitant daily temozolomide has better outcomes in median OS in comparison to radiotherapy alone, 14.6% vs. 12.1%, respectively [40]. Other treatment options include further surgical resection, reirradiation, systemic therapies such as lomustine or bevacizumab [41], combined approaches, or palliative care. Previous studies have indicated that gross total resection improves survival outcomes, independently of molecular status [42]. Unfortunately, supramarginal resection may lead to serious complications [6]. To enable safe glioblastoma resection, variable preoperative and intraoperative surgical technologies, which

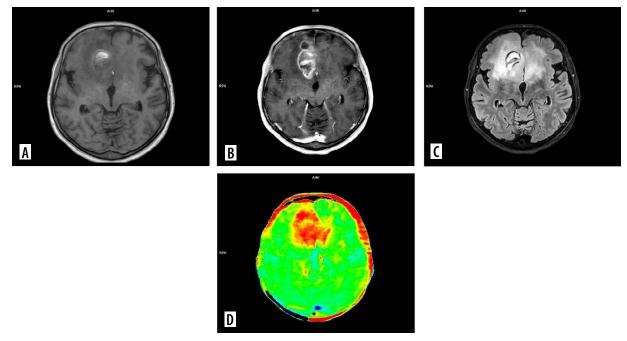


Figure 4. MRI scans of a patient with glioblastoma: A) T1-weighted imaging (T1), B) T1 contrast-enhanced, C) fluid-attenuated inversion recovery (FLAIR), D) chemical exchange saturation transfer (CEST)

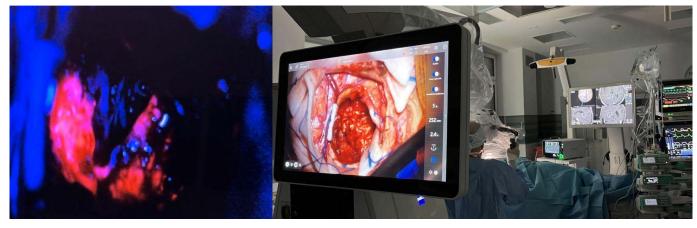


Figure 5. Integrated radiological assessment with intraoperative 5-ALA in the surgical cavity (with the consent of the Department of Neurosurgery, PIM MSWiA)

minimize surgical morbidity, have been developed. Neuroimaging is the first and the most important step in the preoperative approach, especially in the case of involving eloquent brain regions, which could be assessed in DTI. The invasive treatment is based on fluorescence-guided brain surgery. Intraoperatively, infiltrative glioma cells are defined by fluorescent agents such as 5-aminolevulinic acid (5-ALA). This oral chemical agent has been approved by the US Food and Drug Administration for use in neurosurgery. As a result of selective accumulation of agents within the pathological tissue, the tumor borders become fluorescent under blue-light microscopy (Figure 5). This method provides real-time guidance to the surgeon, allowing the resection to be extended toward or beyond the contrast-enhancement areas on MRI. Moreover, 5-ALA-guided resection improves the overall survival of glioblastoma patients vs non-5-ALA-guided surgery [43-45]. Other tools, such as neuronavigation, awake craniotomy, and intraoperative neurophysiological monitoring, lead to limited risk of postoperative neurological deficits [4].

Radiotherapy plays a crucial role in the treatment of glioblastoma after surgery, aiming to enhance local disease control and extend patient survival [46]. The relevant limitation of radiation is necrosis of critical brain structures, depending on the volume of the tissue irradiated and the dose [36]. Patients with glioblastoma frequently exhibit resistance to radiotherapy, which is the major reason for the high mortality rate in this group of patients.

Novel therapeutic strategies are sought to improve the prognosis. Immunotherapeutic approaches, such as immune checkpoint inhibitors, aim to bolster the immune system response against tumor cells. Analyses of immunomodulatory mechanisms in the glioblastoma microenvironment are crucial for designing immunotherapeutic strategies. Thus the PTZ could be a potential target for novel therapeutic options. Immunotherapy and conventional treatment may achieve superior therapeutic outcomes. However, specific biological factors, such as the blood-brain barrier and the unique tumor and immune microenvironment, represent significant challenges in developing novel therapies [40,47].

Conclusions

This review highlighted the role of the PTZ in the preoperative planning of tumor resection and the potential to improve the prognosis of glioblastoma patients. Better imaging characterization of the PTZ could help to develop a biological map of glioblastoma and brain tissue suspected of tumor infiltration to guide neurosurgical resection or radiotherapy planning. Multimodal neurosurgical techniques with better knowledge of the PTZ could determine the optimal resection line while preserving the functional integrity of critical brain structures. Moreover, analysis of the PTZ has the potential to select patients who will benefit from a supramarginal resection. On the other hand, advanced imaging tools have the potential to identify the biological composition of the PTZ and might lead to novel target therapies capable of reducing tumor mass. The differences in immune status across TZ and PTZ provide a potential target for novel immunotherapeutic strategies. More studies are needed to understand the mechanism of glioblastoma recurrence; however, the abnormalities in the PTZ seem to be the baseline of this complicated process.

Disclosures

- 1. Institutional review board statement: Not applicable.
- 2. Assistance with the article: None.
- 3. Financial support and sponsorship: None.
- 4. Conflicts of interest: None.

References

- Wen PY, Weller M, Lee EQ, Alexander BM, Barnholtz-Sloan JS, Barthel FP, et al. Glioblastoma in adults: a Society for Neuro-Oncology (SNO) and European Society of Neuro-Oncology (EANO) consensus review on current management and future directions. Neuro Oncol 2020; 22: 1073-1113.
- Ostrom QT, Cioffi G, Gittleman H, Patil N, Waite K, Kruchko C, Barnholtz-Sloan JS. CBTRUS statistical report: Primary brain and other central nervous system tumors diagnosed in the United States in 2012-2016. Neurooncology 2019; 21 (Suppl 5): v1-v100. DOI: https://doi.org/10.1093/neuonc/noz150.
- Schaff LR, Mellinghoff IK. Glioblastoma and other primary brain malignancies in adults: a review. JAMA 2023; 329: 574-587.
- Luo C, Song K, Wu S, Hameed NUF, Kudulaiti N, Xu H, et al. The prognosis of glioblastoma: a large, multifactorial study. Br J Neurosurg 2021; 35: 555-561.
- Zakharova NE, Batalov AI, Pogosbekian EL, Chekhonin IV, Goryaynov SA, Bykanov AE, et al. Perifocal zone of brain gliomas: application of diffusion kurtosis and perfusion MRI values for tumor invasion border determination. Cancers (Basel) 2023; 15: 2760. DOI: 10.3390/ cancers15102760.
- Giambra M, Di Cristofori A, Valtorta S, Manfrellotti R, Bigiogera V, Basso G, et al. The peritumoral brain zone in glioblastoma: where we are and where we are going. J Sci Res 2023; 101: 199-216.
- Gill BJ, Pisapia DJ, Malone HR, Goldstein H, Lei L, Sonabend A, et al. MRI-localized biopsies reveal subtype-specific differences in molecular and cellular composition at the margins of glioblastoma. Proc Natl Acad Sci U S A 2014; 111: 12550-12555.
- Yan JL, Li C, van der Hoorn A, Boonzaier NR, Matys T, Price SJ. A neural network approach to identify the peritumoral invasive areas in glioblastoma patients by using MR radiomics. Sci Rep 2020; 10: 9748. DOI: 10.1038/s41598-020-66691-6. Erratum in: Sci Rep 2020; 10: 13808.
- Malik N, Geraghty B, Dasgupta A, Maralani PJ, Sandhu M, Detsky J, et al. MRI radiomics to differentiate between low-grade glioma and glioblastoma peritumoral region. J Neurooncol 2021; 155: 181-191.
- Lemée JM, Clavreul A, Aubry M, Com E, de Tayrac M, Eliat PA, et al. Characterizing the peritumoral brain zone in glioblastoma: a multidisciplinary analysis. J Neurooncol 2015; 122: 53-61.
- Śledzińska P, Bebyn M, Szczerba E, Furtak J, Harat M, Olszewska N, et al. Glioma 2021 WHO classification: the superiority of NGS over IHC in routine diagnostics. Mol Diagn Ther 2022; 26: 699-713.
- Singh RR. Next-generation sequencing in high-sensitive detection of mutations in tumors: challenges, advances, and applications. J Mol Diagn 2020; 22: 994-1007.
- Rahimi Koshkaki H, Minasi S, Ugolini A, Trevisi G, Napoletano C, Zizzari IG, et al. Immunohistochemical characterization of immune infiltrate in tumor microenvironment of glioblastoma. J Pers Med 2020; 10: 112. DOI: https://doi.org/10.3390/jpm10030112.
- Walecki J, Chojnacka E, Diagnostyka obrazowa guzów wewnątrzczaszkowych. Onkologia w Praktyce Klinicznej 2007; 3: 177-197.
- Yu H, Lou H, Zou T, Wang X, Jiang S, Huang Z, et al. Applying protein-based amide proton transfer MR imaging to distinguish solitary brain metastases from glioblastoma. Eur Radiol 2017; 27: 4516-4524.

- 16. Inoue A, Ohnishi T, Kohno S, Ohue S, Nishikawa M, Suehiro S, et al. Met-PET uptake index for total tumor resection: identification of 11C-methionine uptake index as a goal for total tumor resection including infiltrating tumor cells in glioblastoma. Neurosurg Rev 2021; 44: 587-597.
- 17. Sebök M, van Niftrik CHB, Muscas G, Pangalu A, Seystahl K, Weller M, et al. Hypermetabolism and impaired cerebrovascular reactivity beyond the standard MRI-identified tumor border indicate diffuse glioma extended tissue infiltration. Neurooncol Adv 2021; 3: vdab048. DOI: 10.1093/noajnl/vdab048.
- Colopi A, Fuda S, Santi S, Onorato A, Cesarini V, Salvati M, et al. Impact of age and gender on glioblastoma onset, progression, and management. Mech Ageing Dev 2023; 211: 111801. DOI: 10.1016/j.mad. 2023.111801.
- 19. Sokolov E, Dietrich J, Cole AJ. The complexities underlying epilepsy in people with glioblastoma. Lancet Neurol 2023; 22: 505-516.
- Gonçalves FG, Chawla S, Mohan S. Emerging MRI techniques to redefine treatment response in patients with glioblastoma. J Magn Reson Imaging 2020; 52: 978-997.
- Henssen D, Meijer F, Verburg FA, Smits M. Challenges and opportunities for advanced neuroimaging of glioblastoma. Br J Radiol 2023; 96: 20211232. DOI: 10.1259/bjr.20211232.
- 22. Li M, Huang W, Chen H, Jiang H, Yang C, Shen S, et al. T2/FLAIR abnormity could be the sign of glioblastoma dissemination. Front Neurol 2022; 13: 819216. DOI: https://doi.org/10.3389/fneur.2022. 819216.
- 23. Csutak C, Ştefan PA, Lenghel LM, Moroşanu CO, Lupean RA, Şimonca L, et al. Differentiating high-grade gliomas from brain metastases at magnetic resonance: the role of texture analysis of the peritumoral zone. Brain Sci 2020; 10: 638. DOI: 10.3390/brainsci10090638.
- 24. D'Alessio A, Proietti G, Sica G, Scicchitano BM. Pathological and molecular features of glioblastoma and its peritumoral tissue. Cancers (Basel) 2019; 11: 469. DOI: 10.3390/cancers11040469.
- 25. Kubben PL, Wesseling P, Lammens M, Schijns OE, Ter Laak-Poort MP, van Overbeeke JJ, van Santbrink H. Correlation between contrast enhancement on intraoperative magnetic resonance imaging and histopathology in glioblastoma. Surg Neurol Int 2012; 3: 158. DOI: 10.4103/2152-7806.105097.
- Gonçalves FG, Chawla S, Mohan S. Emerging MRI techniques to redefine treatment response in patients with glioblastoma. J Magn Reson Imaging 2020; 52: 978-997.
- 27. Norris CD, Quick SE, Parker JG, Koontz NA. Diffusion MR imaging in the head and neck: principles and applications. Neuroimaging Clin N Am 2020; 30: 261-282.
- Sidibe I, Tensaouti F, Roques M, Cohen-Jonathan-Moyal E, Laprie A. Pseudoprogression in glioblastoma: role of metabolic and functional MRI – systematic review. Biomedicines 2022; 10: 285. DOI: https:// doi.org/10.3390/biomedicines10020285.
- Czernicki Z, Horsztyński D, Jankowski W, Grieb P, Walecki J. Malignancy of brain tumors evaluated by proton magnetic resonance spectroscopy (1H-MRS) in vitro. Acta Neurochir Suppl 2000; 76: 17-20.
- 30. Booth TC, Wiegers EC, Warnert EAH, Schmainda KM, Riemer F, Nechifor RE, et al. High-grade glioma treatment response monitoring biomarkers: a position statement on the evidence supporting the use

of advanced MRI techniques in the clinic, and the latest bench-tobedside developments. Part 2: Spectroscopy, chemical exchange saturation, multiparametric imaging, and radiomics. Front Oncol 2022; 11: 811425. DOI: 10.3389/fonc.2021.811425.

- Weinberg BD, Kuruva M, Shim H, Mullins ME. Clinical applications of magnetic resonance spectroscopy in brain tumors: from diagnosis to treatment. Radiol Clin North Am 2021; 59: 349-362.
- 32. Koike H, Morikawa M, Ishimaru H, Ideguchi R, Uetani M, Miyoshi M. Amide proton transfer-chemical exchange saturation transfer imaging of intracranial brain tumors and tumor-like lesions: our experience and a review. Diagnostics (Basel) 2023; 13: 914. DOI: 10.3390/ diagnostics13050914.
- 33. Akbari H, Kazerooni AF, Ware JB, Mamourian E, Anderson H, Guiry S, et al. Quantification of tumor microenvironment acidity in glioblastoma using principal component analysis of dynamic susceptibility contrast enhanced MR imaging. Sci Rep 2021; 11: 15011. DOI: 10.1038/s41598-021-94560-3.
- 34. Neal A, Moffat BA, Stein JM, Nanga RPR, Desmond P, Shinohara RT, et al. Glutamate weighted imaging contrast in gliomas with 7 Tesla magnetic resonance imaging. NeuroImage Clin 2019; 22: 101694. DOI: 10.1016/j.nicl.2019.101694.
- 35. Kumar M, Nanga RPR, Verma G, Wilson N, Brisset JC, Nath K, Chawla S. Emerging MR imaging and spectroscopic methods to study brain tumor metabolism. Front Neurol 2022; 13: 789355. DOI: https://doi.org/10.3389/fneur.2022.789355.
- Kwiatkowska-Miernik A, Mruk B, Sklinda K, Zaczyński A, Walecki J. Radiomics in the diagnosis of glioblastoma. Pol J Radiol 2023; 88: 461-466.
- 37. Prasanna P, Patel J, Partovi S, Madabhushi A, Tiwari P. Radiomic features from the peritumoral brain parenchyma on treatment-naïve multi-parametric MR imaging predict long versus short-term survival in glioblastoma multiforme: preliminary findings [published correction appears in Eur Radiol 2017]. Eur Radiol 2017; 27: 4188-4197.
- Long H, Zhang P, Bi Y, et al. MRI radiomic features of peritumoral edema may predict the recurrence sites of glioblastoma multiforme. Front Oncol 2023; 12: 1042498. DOI: 10.3389/fonc.2022.1042498.

- Gritsch S, Batchelor TT, Gonzalez Castro LN. Diagnostic, therapeutic, and prognostic implications of the 2021 World Health Organization classification of tumors of the central nervous system. Cancer 2022; 128: 47-58.
- 40. Tan AC, Ashley DM, López GY, Malinzak M, Friedman HS, Khasraw M. Management of glioblastoma: state of the art and future directions. CA Cancer J Clin 2020; 70: 299-312.
- 41. Pinkiewicz M, Pinkiewicz M, Walecki J, Zawadzki M. A systematic review on intra-arterial cerebral infusions of chemotherapeutics in the treatment of glioblastoma multiforme: the state-of the-art. Front Oncol 2022; 12: 950167. DOI: 10.3389/fonc.2022.950167.
- 42. Brown TJ, Brennan MC, Li M, Church EW, Brandmeir NJ, Rakszawski KL, et al. Association of the extent of resection with survival in glioblastoma: a systematic review and meta-analysis. JAMA Oncol 2016; 2: 1460-1469.
- 43. Zeppa P, De Marco R, Monticelli M, Massara A, Bianconi A, Di Perna G, et al. Fluorescence-guided surgery in glioblastoma: 5-ALA, SF or both? Differences between fluorescent dyes in 99 consecutive cases. Brain Sci 2022; 12: 555. DOI: 10.3390/brainsci12050555.
- 44. Baig Mirza A, Christodoulides I, Lavrador JP, Giamouriadis A, Vastani A, Boardman T, et al. 5-Aminolevulinic acid-guided resection improves the overall survival of patients with glioblastoma – a comparative cohort study of 343 patients. Neurooncol Adv 2021; 3: vdab047. DOI: 10.1093/noajnl/vdab047.
- 45. Hadjipanayis CG, Widhalm G, Stummer W. What is the surgical benefit of utilizing 5-aminolevulinic acid for fluorescence-guided surgery of malignant gliomas? Neurosurgery 2015; 77: 663-673.
- 46. Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJ, et al.; European Organisation for Research and Treatment of Cancer Brain Tumor and Radiotherapy Groups; National Cancer Institute of Canada Clinical Trials Group. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. N Engl J Med 2005; 352: 987-996.
- 47. Czarnywojtek A, Borowska M, Dyrka K, Van Gool S, Sawicka-Gutaj N, Moskal J, et al. Glioblastoma multiforme: the latest diagnostics and treatment techniques. Pharmacology 2023; 108: 423-431.